

# Upstream BioProcessing Questionnaire

Please answer the following questions as completely as possible.

## I. Customer Information

CONTACT PERSON

COMPANY NAME

DESIGNATION

CONTACT NUMBER

DEPARTMENT

EMAIL ADDRESS

## II. General Details

### 1. Target Product

- |   |   |
|---|---|
| <input type="checkbox"/> Secreted Protein     | <input type="checkbox"/> Virus Production     |
| <input type="checkbox"/> Non secreted protein | <input type="radio"/> Human                   |
| <input type="checkbox"/> Cell bank            | <input type="radio"/> Veterinary              |
| <input type="checkbox"/> mAbs                 | <input type="checkbox"/> Cultivated Meat      |
| <input type="checkbox"/> Cell Therapy         | <input type="checkbox"/> <input type="text"/> |

### 2. Cell Type

- Adherent cell (Proceed to Adherent Cell Questionnaire)
- Suspension cell (Proceed to Suspension Cell Questionnaire)

### 3. What process development (PD)/optimization do you require?

- Cell line development
- Upstream development  
*e.g. bioreactor media optimization, harvest protocol*
- Downstream development  
*e.g. optimization of platform DS process*
- No PD required. Process to be transferred at existing scale to manufacturing.

4. Do you require any of the following? Please attach an extra sheet if additional services are required.

- Analytical Method Validation
- cGMP manufacturing and lot release
- Stability testing
- Sterility testing of final product
- Adventitious virus testing
- 

## Adherent Cells Questionnaire

### III. Experiment Details

#### a. General Details

1. Cell Line

- CHO
- MDCK
- 
- Vero
- HEK 293
- Hybridoma
- SF-9

2. Any special features or peculiarities of the cell line or methods

3. Intended Use

- Human Use
- Animal Use

4. Current Culture System

- T-flask
- Roller bottle
- Spinner flask
- Cell factory
- Cell stack
- Hyper flask
- Stirred Tank Bioreactor with Carriers
- 

Indicate Capacity in liters (L):

5. If carriers are used, please specify type and amount of carrier.

- Microbeads, Specify:
- Fibrous matrices, Specify:
- Others, Specify:

Amount of carriers:  grams

6. Culture condition for cell growth

- Media
- Serum
- Temperature

7. Currently using serum-free culture medium?

- Yes
- No

8. Concentration of additives

- Sodium Bicarbonate     Others
- Hepes Buffer

9. Cell Harvesting Required

- Yes
- No

10. Use of trypsin during cell harvest

- Yes                       Use others. Please specify:
- No

11. Cell Quantification

- Manual counting                       Nuclei counting
- Auto-counter                               Others

12. Access to a bio-analyzer for measuring glucose, lactate, glutamine, etc.

- Yes
- No

13. System preference

- Single-Use Preference                       No Preference
- Multiple-Use Preference

14. Scale-up plan in terms of number of cells

10<sup>9</sup>  
 10<sup>10</sup>

10<sup>11</sup>  
 10<sup>12</sup>

>10<sup>13</sup>

15. Scale-up plan in terms of volume

50L  
 100L

500L  
 Others

## b. Protein Production

1. Culture period prior to harvesting

3 Days  
 5 Days  
 7 Days

Other (Please Specify): days

2. Protein extraction method

By cell harvest  
 By medium harvest

Freeze/Thaw method  
 Others

## c. Cell Therapy

1. Target

Autologous Cell Therapy  
 Allogeneic Cell Therapy

Research Use  
 Others

2. Cell Source

Bone Marrow  
 Adipose-derived  
 iPS  
 Embryo

Placenta  
 Umbilical  
 Dermal fibroblast  
 Others

### III. Experiment Details

#### d. Virus Production

1. Virus Type/Strain

Secreted Virus

Non-secreted Virus Virus

Strain:

2. Please describe the Virus Strain? (ds, ssDNA, ds, +/- ssRNA, enveloped, nonenveloped, temperature sensitivity, etc.)

3. Cell density prior to infection in current culture system

4. Multiplicity of Infection (MOI)

5. Period of time for cell lysis to occur after infection in current culture system

6. Culture condition post infection

Media

Serum

Temperature

7. Best phase for infection

Right after seeding

Plateau phase

Exponential phase

8. Is the virus stable during post infection?

9. Virus titer in current culture system (dose/ml)

10. Best time to harvest the virus

11. Is there CPE (cytopathic effect) after infection? When?

- Yes;
- No

12. What kind of CPE is formed (e.g. syncytium, destruction, etc.)

13. Cell lysis post infection

- Yes;
- No

14. Number of harvests that could be done during post infection period

- Single Harvest
- Multi-harvest for  times
- Continuous Harvest for  days

15. Do cells keep propagating after virus infection?

- Yes. Indicate fold increase post infection:
- No

16. For virus production, annual manufactured dose

# Fermentation/Suspension Cells

## III. Experiment Details

### a. General Details

#### 1. Application

- Microbial Culture                      Adherent Cell Culture  
 Suspension Mammalian Culture

#### 2. Cells Culture

- Bacteria                       Cell Line:   
 Yeast                       Others   
 Fungi

#### 3. For mammalian cells, specify type and amount of carrier used

- Microbeads, Specify:   
 Fibers, Specify:   
 Others, Specify:   
  
**Amount of carrier:**

#### 4. Product

- Secreted Protein                       Secreted Virus  
 Non-Secreted Protein                       Non-Secreted Virus  
 Fermentation                       Cell Banking  
 Others

#### 5a. Current Culture System

- Spinner Flask                       Others   
 Stirred Tank Bioreactor

#### 5b. Current Culture Scale in liters (L):

#### 6. Current Process Mode

- Batch                       Continuous  
 Fed-Batch                       Others

<p><b>7. Culture condition for cell growth</b></p>	<p><input type="checkbox"/> Media <input type="text"/></p> <p><input type="checkbox"/> Serum <input type="text"/></p> <p><input type="checkbox"/> Temperature <input type="text"/></p>
<p><b>8. Currently using serum-free culture medium?</b></p>	<p><input type="checkbox"/> Yes</p> <p><input type="checkbox"/> No</p>
<p><b>9. Concentration of additives</b></p>	<p><input type="checkbox"/> Sodium Bicarbonate: <input type="text"/></p> <p><input type="checkbox"/> HEPES Buffer: <input type="text"/></p> <p><input type="checkbox"/> Others <input type="text"/></p>
<p><b>10. Cooling system required?</b></p>	<p><input type="checkbox"/> Yes</p> <p><input type="checkbox"/> No</p>
<p><b>11. Temperature sensitive?</b></p>	<p><input type="checkbox"/> Yes</p> <p><input type="checkbox"/> No</p>
<p><b>12. pH Sensitive?</b></p>	<p><input type="checkbox"/> Yes</p> <p><input type="checkbox"/> No</p>
<p><b>13a. Shear Stress Tolerance</b></p>	<p><input type="checkbox"/> High Sensitivity                      <input type="checkbox"/> High tolerance to shear stress</p> <p><input type="checkbox"/> Medium Sensitivity</p>
<p><b>13b. Preferred Impeller Type</b></p>	<p><input type="text"/></p>
<p><b>14. Scale up plan in terms of volume in liters</b></p>	<p><input type="text"/></p>



### III. Experiment Details

#### b. Process Control

##### 1. Reactor Size

Minimum working volume:

Maximum working volume:

##### 2. Agitation Speed

Range:  rpm to  rpm

##### 3. Measurements Required

Temperature  °C to  °C

pH  to

DO  % to  %

Redox  mV to  mV

Turbidity  pCO<sub>2</sub>

Foaming  O<sub>2</sub>/CO<sub>2</sub> in Exhaust Gas

Level  Others

##### 4. Temperature Control

Double wall vessel  Heating Pad

Heating Jacket  Heating/Cooling Pad

Others

##### 5. pH Control

Addition of Base  Addition of CO<sub>2</sub>

Addition of Acid  Others

##### 6. Dissolved Oxygen Control

Impeller Speed  Gas Flow Rate

Addition of O<sub>2</sub>  Others

##### 7. Foaming

High  Not yet determined

Low

**8. Applied Gases for Aeration**

- Air
  - Air + O<sub>2</sub>
  - Air + O<sub>2</sub> + N<sub>2</sub>
  - Air + O<sub>2</sub> + N<sub>2</sub> + CO<sub>2</sub>
- Others
- Mixing System:**  
 Yes  No

**9. Airflow**

- Range:  vvm to  vvm
- Control:
- Regulator (manual)
  - Mass Flow Controller
  - Others

**10. Aeration Delivery**

- Sparger, Type:
- Ring Sparger
  - Microsparger
  -
- Overlay
- Both

**11. Pressure Control Requirements**

**12. Other Special Requirements**

**Important:** Save the completed PDF form (use menu File - Save).